

Nature. 2005 December 1; 438(7068): 633–638.

2B6P, 2B6O, 1YMG

Extracellular AQP0

H1 – blue

H2 – light blue

HB - cyan

H3 – light green

H4 – green

H5 – yellow

HE - gold

H6 – orange

CT – red

Cytoplasm

Figure 7: Lipids surrounding the AQP0 tetramer mediate the crystal contacts. Horizontal section through an AQP0 tetramer (blue) showing one of the water molecules in the pore (red) and the tightly packed acyl chains of the surrounding shell of lipid molecules (yellow).

Supplementary 2B6P pH=6.5 Table1: Crystallographic statistics of non junctional AQP0 (X-ray).

None of the AQP 3D crystals examined so far contain lipids, and 2D crystals of AQPs can form with a variety of different lipids, suggesting that AQPs have neither a requirement for specific lipids nor high-affinity lipid binding sites hydrophobic. Nevertheless, our density map revealed that between the AQP0 tetramers are horseshoe-shaped features characteristic of lipid molecules (Fig. 4a). Indeed, close inspection revealed that lipids bridge-hydrophobic all the contacts between tetramers within a layer and that the tetramers have essentially hydrophobic lateral interaction. In composite omit maps, we could identify nine lipids per AQP0 monomer, which we modeled as complete or partial molecules of dimyristoyl phosphatidyl choline (DMPC, the lipid used for 2D crystallization) (Fig. 4a). Phospholipid headgroups have a chiral center at C2 of the glycerol, and the DMPC we used is a racemic mixture. Density is weak or absent at most C2 positions in our map, and often at the attached ester group as well, suggesting that there is little or no selectivity for the biological enantiomer. Very strong density for the phosphate groups, weaker but well defined density for the trimethyl amine groups of the cholines, and unambiguous density for the acyl chains allowed us to build and refine a model in which we chose an enantiomer for each lipid more or less arbitrarily. We have not yet attempted to refine the two alternatives with 50% occupancy each. We have annotated these lipids as PC1 to PC9 (Fig. 4b; Suppl. Fig. 6). PC1 to PC7 have extensive protein contacts and appear to represent “annular lipids” immediately adjacent to a membrane embedded protein. PC8 and PC9 are not in contact with protein and thus represent bulk lipids. A detailed description of protein-lipid contacts is provided in Supplementary Materials. As AQP0 has no tight lipid binding sites, interactions between the annular lipids and the AQP0 subunits are likely to represent the kind of contacts that occur between any membrane protein and the lipids surrounding it.

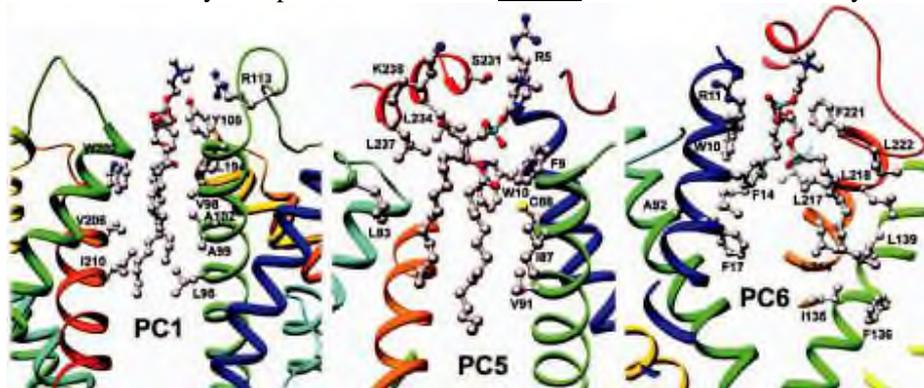


Figure 4: Lipid-protein interactions in double-layered AQP0 2D crystals. a. Vertical slab through the 2Fo-Fc density map with modelled lipid molecules, revealing the two lipid bilayers in the double-layered AQP0 2D crystal. b. The nine lipids surrounding an AQP0 monomer in the 2D crystal. Lipids PC1 to PC7 are annular lipids, whereas lipids PC8 and PC9 are bulk lipids with no direct protein contacts. See Supplementary Figure 6 for a stereo view. c – e. Three examples of lipids sandwiched in

between two AQP0 molecules. The acyl chains of PC1 adopt a closed (c), those of PC5 a slightly splayed (d), and those of PC6 a widely splayed conformation (e).

Annular lipids must adapt to the irregular surface of a transmembrane protein to create a smooth interface for bulk lipids. This fit limits the mobility (and perhaps the chemistry) of annular lipids, as their conformations are partially defined by the

protein surface. In our 2D arrays, most of the **annular lipids** are sandwiched between **two tetramers** and thus mediate **lattice interactions** (Suppl. Fig. 7). This packing further restricts their **conformations**. The **cell dimensions** of our reconstituted **junctions** are the same as those in **thin junctions** between **lens fibre cells**²⁸. We therefore suggest that the **lipid-protein interactions** we observe in our 2D crystals with the artificial **lipid DMPC** are representative of those formed by **AQP0 tetramers** with native **lipids** in **lens fibre cell membranes**.

The **lipids** form a one-molecule wide **annular shell** around the protein. The positions of the **headgroups** vary by only ± 2 Å in the direction perpendicular to the **membrane plane**, with a separation of about **34 Å** from **phosphate to phosphate**. The dimensions of the **bilayer** correspond **closely** to those of fully hydrated, fluid phase **DMPC**²⁹. A hydrated network of **hydrogen bonds** and **salt bridges** holds the **lipid phosphates** in place. Protein **groups interacting with phosphates** include **three Arginine side chains**, a **tyrosine hydroxyl** that mediates one of the **Arginine contacts**, a **lysine**, a **tryptophan indole nitrogen**, a **glutamine side-chain amide**, and at least one **main-chain amide**. Similar **interactions** have been described for specifically bound **lipids**³⁰.

Acyl chains fill the gaps between adjacent **tetramers**. Their **conformations** clearly adapt to the knobs and grooves of the apposed **hydrophobic** protein surfaces. **Figures 4c-e** illustrate **three examples**. **PC1** in the **extracellular leaflet** is the best ordered of the **nine DMPC molecules**. Its **acyl chains** are nearly fully extended, packed against those of **PC2** and **PC3** and sandwiched between five **non-polar side chains** from one **AQP0** and **three** from the other. **PC5** in the **cytoplasmic leaflet** has somewhat less extended **acyl chains**. The **phosphate** receives a **hydrogen bond** from the **indole nitrogen** of **Trp10** and **Lys238** (as well as the poorly ordered **N-terminal** segment) of an adjacent subunit. The **acyl chains**, packed between those of **PC4** and **PC6**, **contact** four **hydrophobic side chains** from one subunit (including the **hydrophobic face** of **Trp10**) and **three** from another. **PC6**, also in the **cytoplasmic leaflet**, has widely splayed **acyl chains**, separated by side chains from the **two** apposed **AQP0 molecules**. **Phe14** of one molecule and **Leu217** of another are in van der Waals **contact** through the gap: the only direct **interaction** between **tetramers** **within** a layer.

PC8 and **PC9** lie near the fourfold axis. They do not **contact** protein and thus represent bulk **lipids**. Neither is as well ordered as the **annular lipids**. Indeed, **PC8** (in the **cytoplasmic leaflet**) is probably only statistically ordered (**two**, rather than **four**, **molecules** about a fourfold), as there is space for only one of the **two acyl chains** and no density for the **headgroup**. The **headgroup** of **PC9** lies about **3 Å** **closer** to the midplane of the **bilayer** than those of the four other **extracellular leaflet lipids**; the **bilayer thickness** may therefore be influenced by adjacency to the protein. **2B6P pH=6.5 non junctional 2B6O pH=10.5**

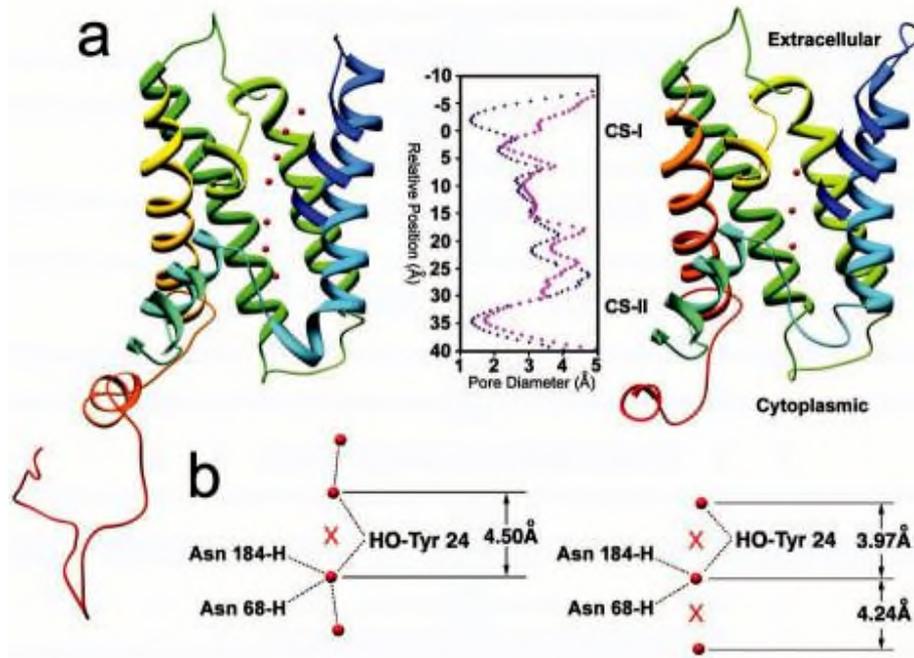


Figure 3: The water pore in AQP0. a. The pore in **non-junctional AQP0** (left) contains **seven water molecules** (red spheres), while the pore in **junctional AQP0** contains only **three water molecules** (right). Calculated pore profiles (middle) corroborate that the pore.

In our initial report of the **closed water pore** in **junctional AQP0**⁷, we proposed that **AQP0** and other **Aquaporins** may be in a dynamic equilibrium between an open and a **closed pore conformation**. We also suggested that **pore closure** may be triggered by the stabilisation of an alternative **conformation** of **Arg187** (part of the **ar/R constriction site**) seen in the structure of **junctional AQP0**. A recent molecular dynamics study supports this notion, as it showed that **Arg189** in **AQPZ** (corresponding to **Arg187** in **AQP0**) could adopt **two conformations**²⁰. The “UP” state, which is

seen in most **AQP** crystal structures, had an open pore, filled by a continuous single file of **water**. The “DOWN” state, seen in our structure of **junctional AQP0**, had a pore completely blocked by the **Arg** side chain, and prolonged blockage resulted in loss of all **water molecules** from the pore. While attractive, a **conformational switch** of the **Arginine** in the **ar/R constriction site** cannot be the only mechanism for **AQP** gating, because **Arg187** is in the “DOWN” state not only in our **closed, junctional AQP0** but also in the open, **non-junctional AQP0** structure⁸. The main difference between the open and **closed pore** lies in the **conformation** of the side chain of **Met176** (see above), a residue not present in **AQPZ**.

The distances between the **three water molecules** ($\geq 4 \text{ \AA}$) in the **closed pore** are too long for **hydrogen bonding** (Fig. 3b, right; Suppl. Fig. 5, right). The **water** coordinated to the Asn residues of the **two NPA motifs** donates a **hydrogen bond** to the hydroxyl **group** of **Tyr24**, which in turn donates a **hydrogen bond** to the **water** molecule in the **extracellular half** of the **water pathway** (Fig. 3b, right; Suppl. Fig. 5, right). The corresponding **two water molecules** in the **open water pore** of **non-junctional AQP0** have the same **hydrogen bonding** pattern (Fig. 3b, left; Suppl. Fig. 5, left), and all the other **water molecules** are in **hydrogen-bonding** distance to each other. “**Phenolic barrier** created by **Tyr24**, a residue not seen in the other known **AQP** structures, may be responsible for the poor **water conductance** of **AQP0** as compared to other **AQPs**, which contain a continuous line of **hydrogen-bonded water molecules**. The space occupied by **Tyr24** may also explain why the open **AQP0 pore** contains only **seven water molecules** while molecular dynamics studies showed **eight waters** in **AQP1**^{21,22} and **AQPZ**²⁰ and **nine** in **GlpF**²³.

AQP0 water conductance is **pH-dependent** with a maximum at **pH 6.5** and only about half the activity at **pH 10.5**¹². These **conductance** characteristics are not changed by proteolytic cleavage of **AQP0**²⁴. As our structure, obtained with the double-layered 2D crystals grown at **pH 6**⁷, reveals fewer **water molecules** in the **pore** than the structure determined from the 3D crystals grown at **pH 10.5**⁸, **pore closure** appears to be a result of **junction** formation, not **pH** shift.

OPEN 2B6P pH=6.5 non junctional															
REMARK	525	HOH	A	328	DISTANCE =	5.22	ANGSTROMS		2B6P						
REMARK	525	HOH	A	340	DISTANCE =	6.88	ANGSTROMS								
REMARK	525	HOH	A	347	DISTANCE =	6.00	ANGSTROMS								
REMARK	525	HOH	A	348	DISTANCE =	5.04	ANGSTROMS								
REMARK	525	HOH	A	362	DISTANCE =	5.54	ANGSTROMS								
REMARK	525	HOH	A	376	DISTANCE =	5.45	ANGSTROMS								
HELIX	1	1	PHE	A	9	LEU	A	32	1	24					
HELIX	2	2	GLY	A	37	GLY	A	64	1	28					
HELIX	3	3	ASN	A	68	GLY	A	78	1	11					
HELIX	4	4	SER	A	82	THR	A	108	1	27					
HELIX	5	5	SER	A	126	ASP	A	150	1	25					
HELIX	6	6	SER	A	159	GLY	A	180	1	22					
HELIX	7	7	ASN	A	184	ARG	A	196	1	13					
HELIX	8	8	TRP	A	202	PHE	A	221	1	20					
HELIX	9	9	SER	A	229	LEU	A	237	1	9					
junctional 2B6O pH=10.5 half OPEN															
REMARK	525	HOH	A	281	DISTANCE =	7.45	ANGSTROMS		2B6O						
REMARK	525	HOH	A	282	DISTANCE =	5.39	ANGSTROMS								
REMARK	525	HOH	A	305	DISTANCE =	7.20	ANGSTROMS								
REMARK	525	HOH	A	314	DISTANCE =	5.12	ANGSTROMS								
REMARK	525	HOH	A	346	DISTANCE =	9.27	ANGSTROMS								
HELIX	1	1	ARG	A	5	LEU	A	32	1	28					
HELIX	2	2	LEU	A	39	VAL	A	59	1	21					
HELIX	3	3	GLY	A	60	ILE	A	62	5	3					
HELIX	4	4	ASN	A	68	GLY	A	78	1	11					
HELIX	5	5	SER	A	82	THR	A	108	1	27					
HELIX	6	6	SER	A	126	TYR	A	149	1	24					
HELIX	7	7	SER	A	159	MET	A	176	1	18					
HELIX	8	8	ASN	A	184	ARG	A	196	1	13					
HELIX	9	9	TRP	A	202	PHE	A	221	1	20					
HELIX	10	10	SER	A	229	LEU	A	234	1	6					
HELIX	11	11	SER	A	235	LEU	A	237	5	3					
SITE	1	AC1	6	ARG	A	196	MC3	A	269	MC3	A	270	MC3	A	272
SITE	2	AC1	6	HOH	A	273	HOH	A	304						
SITE	1	AC2	8	ALA	A	102	VAL	A	103	TYR	A	105	SER	A	106
SITE	2	AC2	8	MC3	A	270	HOH	A	291	HOH	A	301	HOH	A	327
SITE	1	AC3	9	LEU	A	83	LEU	A	84	ILE	A	87	VAL	A	90
SITE	2	AC3	9	VAL	A	91	LEU	A	94	LYS	A	238	MC3	A	267
SITE	3	AC3	9	MC3	A	271									
SITE	1	AC4	8	ARG	A	5	SER	A	6	PHE	A	9	TRP	A	10
SITE	2	AC4	8	LEU	A	84	CYS	A	88	MC3	A	266	MC3	A	272
SITE	1	AC5	4	ALA	A	7	TRP	A	10	ARG	A	11	PHE	A	14
SITE	1	AC6	2	MC3	A	264	MC3	A	272						
SITE	1	AC7	8	LEU	A	95	TYR	A	105	ILE	A	193	LEU	A	194
SITE	2	AC7	8	ARG	A	196	MC3	A	264	MC3	A	265	HOH	A	326
SITE	1	AC8	1	MC3	A	266									
SITE	1	AC9	3	MC3	A	264	MC3	A	267	MC3	A	269			